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CONVENIENT c-3 OXIDATION OF DEOXY AND AMINODEOXY SUGARS*

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In spite of numerous reports, $1,3$ the simple and efficient oxidation of primary or secondary hydroxyl groups of protected sugar derivatives to chiral synthons offering versatile possibilities for further transformations, is still a chalenging synthetic problem. Arrick and coworkers² successfully oxidized the primary or exocyclic secondary hydroxyl groups of various carbohydrate derivatives with the chromium trioxidepyridine complex.However the influence of the steric orientation of the hydroxyl function to be oxidized (axial or equapyridine complex.However the influence of the steric orienta-
tion of the hydroxyl function to be oxidized (<u>axial</u> or <u>equa-</u>
torial in hexopyranosides and <u>exo</u> or <u>endo</u> in five membered rings) on the yield was recognized, together with the formation of varying quantities of by-products. In addition, a failure in analogous oxidations of isolated endocyclic hydroxyl groups attached to the ring-carbons of furanoid and pyranoid systems has been reported². In contrast, Garegg and Samuelsson³ showed that the addition of acetic anhydride as promoter to the in situ generated chromium trioxide-pyridine complex in dichloromethane resulted in the rapid oxidation (10-15 min) of primary or secondary alcohol functions of carbohydrates to **Q1990 by Organic Preparations and Procedures Inc.**

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carbonyl groups, even in cases when other oxidation methods failed. Our study of the syntheses and transformations of deoxy and aminodeoxy sugars led us to extend the Garegg- $-$ Samuelsson procedure³ to protected 2-deoxy- and 2-amino-2deoxy sugar derivatives. The C-3 ulose compounds produced have turned out to be invaluable chiral synthons⁴ for various synthetic conversions, including inversion of the configuration at C-3 and generation of axially- or equatoriallyoriented amino functions; thus offers a simple access to 3-amino-2,3-dideoxy- and **2,3-diamino-2,3-dideoxyhexoses,** as well as to C-3-branched carbohydrate derivatives etc.⁵

The starting materials, methyl **4,6-0-benzylidene-2-deoxy-** β -D-arabino-hexopyranoside (la)⁶ and the corresponding α -D-<u>ribo</u>-analogue (<u>lb</u>)⁷, methyl 2-benzamido-4,6-0-benzylidene-2-
deoxy-∝-D-glucopyranoside (<u>lc</u>)⁸ and its 2-benzyloxycarbonyl derivative (<u>ld</u>)⁹, methyl 2-benzamido-4,6-0-benzylidene-2-
deoxy-^α-Q-allopyranoside (<u>le</u>)⁸ and methyl 2-benzamido-4,6-0deoxy- α -<u>D</u>-allopyranoside (<u>le</u>)⁸ and methyl 2-benzamido-4,6-0-
benzylidene-2-deoxy- α -D-altropyranoside (lf)¹⁰ were prepared according to literature methods. Oxidation of compounds la-f was accomplished with a 1:4 molar ratio of the hydroxyl compound and the chromium trioxide-pyridine-acetic anhydride system in dry dichloromethane as detailed in Table 1 and in the general procedure. Under these conditions, no modification of the protecting groupswas observed.

The ¹H-NMR data of the hexopyranosid-3-uloses obtained (la-f) are summarized in Table 3.

The formation of by-products (such as the corresponding 3-0acetate) was not observed in either of the oxidations. The high yields (88-98 %) of the ulose derivatives 2a-d clearly

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prove that the oxidizing system used is not sensitive to steric factors and that it is useful for $C-3$ equatorial (la,c, **CONVENIENT C-3 OXIDATION OF DEOXY- AND AMINODEOXY SUGARS**
prove that the oxidizing system used is not sensitive to
steric factors and that it is useful for C-3 <u>equatorial</u> (<u>la,c,</u>
d) and <u>axial</u> (<u>lb,e,f</u>) hydroxyl comp tuent at C-2.

TABLE 1. Conversion of l_a-f to Hexopyranosid-3-uloses (2a-d)

Starting material	Product			
$\underline{\underline{\mathsf{la}}}$	2a	80	88	
$\frac{1}{b}$	2b	80	97	
lc	2c	35	98	
$\underline{\underline{\mathbf{1}}}\underline{\underline{\mathbf{d}}}$	2d	45	96	
$\underline{\mathbf{ie}}$	2c	35	96	
\underline{If}	2c	40	94	
				$Time(min)$ $Yield(X)$

TABLE 2. Physical Data for C-3 Uloses (2a-d)

The rate of oxidation is independent of the anomeric configuration, while substitution with an electron-withdrawing moiety

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at C-2 (i.e. in compounds lc-e) seems to shorten the reaction time considerably.

 $Bz : COC_6H_5$

Cbz: $COOCH_2C_6H_5$
In the case of the altropyranoside lf, the oxidation proceeded with the inversion of the configuration of the <u>axial</u> proceeded with the inversion of the configuration of the <u>axial</u>
benzamido group at C-2, to give <u>2c</u> with physical data identical benzamido group at $C-2$, to give $2c$ with physical data identical
with those of the product obtained from $1c$ and $1e$. Similar results have been reported by Baker and Buss⁸ and Ali and Richardson 11 in the oxidation of the corresponding C-2 acetamido or azido derivative, respectively, by the Pfitzner-Moffat reagent or with dimethylsulfoxide-acetic anhydride. It is most probable that the inversion of the axial C-2 benzamido group, probable that the inversion of the <u>axial</u> C-2 benzamido group,
close to the site of oxidation, proceeds <u>via</u> an acid-catalyzed enolization involving a carbanion at C-2.

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	Ketosugars			
Hydrogen	2a	2 _b	2c	$\overline{2d}$
ArH	$7.30 - 7.60$		$7.30 - 7.60$ $7.30 - 7.95$	7.38-7.53
H_{1}	4.78	5.15	5.37	5.22
H ₂	3.39	2.85		
H_2 ,	3.23	2.67	5.15	4.65
$H_{\mathbf{A}}$	4.34	4.32	4.48	4.48
H_5	3.67	4.17	4.15	4.15
H_6	4.48	4.41	4.43	4.38
H_6 ,	3.92	3.92	4.03	3.95
NH			6.95	5.63
$J_{1,2}$	3.0	4.0		
$J_{1,2}$	8.0	1.0	4.4	4.5
$J_{2,2}$	14.5	14.5		
$J_{4,5}$	9.5	10.5	9.5	9.5
$J_{5,6}$	5.0	4.5	4.0	4.1
$J_{5,6}$	10.0	10.0	9.9	9.8
$^{J}6, 6,$	10.5	10.0	9.9	9.8
J_2 , NH			8.0	6.0

TABLE 3. 'H-NMR(6) and Coupling Constans (Hz) **of C-3 Ketosugars**

EXPERIMENTAL SECTION

Melting points were determined with a Kofler hot-stage apparatus and are uncorrected. Specific optical rotafions were measured with a Perkin-Elmer Polarimeter 241. The H-NMR spectra (200 HHz) were recorded with a Bruker WP 200 SY instrument for solutions in CDC13 (TMS internal standarp). Dichloromethane was distilled from P 0 and kept over 3 A molecular sieves. Pyridine was purifieg 5by distillation from P O5 and stored over KOH. Chromium trioxide (Fluka p.a.) was dried over P₂O_E in a vacuum desiccator. Ethyl acetate was freed from acid and **alcohol. Thin layer chromatography was performed on precoated silica gel plates (Merck 60 F**) **with the solvent systems: (A) chloroform; (B) 98:2 chl?&%form-methanol and (C) 7:3 chloroform-acetone. For column chromatography, Kieselgel 40 (Merck) was used.**

General Procedure for Oxidation. For 10 mmol of the sugars

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 $(la-f)$ to be oxidized the chromium trioxide-pyridine complex was prepared by the addition of dry chromium trioxide (4.0 g; 40 mmol) to a mixture of dry dichloromethane (85 ml) and dry pyridine (6.44 ml, 80 mmol) with vigorous stirring. The mixture was stirred at room temperature for 15 min and a concentrated solution of la or lb or a dilute suspension of lc-f in dry dichloromethane, and acetic an'hydride (3.80 ml; 40 mmol) were at room temperature for 15 min and a conce
<u>la</u> or <u>lb</u> or a dilute suspension of <u>lc-f</u> added to the deep-red solution. A tarry deposit formed almost at once and the color of the solution changed to dark-brown. The reaction was monitored by TLC and when all the starting sugar had reacted, the mixture was poured onto the top of a short column filled with silica gel and ethyl acetate, and the product was separated from the precipitated chromium compounds by means of adsorptive filtration with ethyl acetate as the eluent. The colorless eluate was concentrated under diminished pressure at 40⁰ (bath temperature) and the residue was codistillated with toluene to remove traces of acetic acid and pyridine. The products were recrystallized from methanol. The physical data of the hexopyranosid-3-uloses obtained (2a-d) are summarized in Table 2.

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